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Intramolecular conjugate addition of γ - and δ -trichloroacetimidoyloxy- α , β -unsaturated esters in an acyclic system

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Abstract

An intramolecular conjugate addition of γ - and δ -trichloroacetimidoyloxy- α , β -unsaturated esters, a new way to construct 1,2-amino or 1,3-amino alcohol moieties in an acyclic system, is described. Very concise synthesis of D-vancosamine and 3-*epi*-D-vancosamine derivatives was also achieved utilizing this methodology.

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Keywords: Intramolecular conjugate addition; Trichloroacetimidate; Vancosamine

1. Introduction

Amino sugars, especially deoxyamino sugars, are found in clinically important antibiotics such as antimicrobial macrolides and anthracycline antitumor antibiotics.¹ In most instances, the sugar parts of these antibiotics are essential for biological activity; however, the functions of the sugar moieties have not yet been evaluated.² We envisaged that a modification of the sugar moieties of these antibiotics may serve as a tool for investigating the significance of amino sugars and the structure—activity relationship, and elucidating the biosynthetic route of antibiotics.^{3,4} For this purpose, a versatile and synthetic route for deoxyamino sugars is highly desirable. We were especially interested in developing a new synthetic route for deoxyamino sugars from non-sugar materials.^{5,6}

1,2- and 1,3-amino alcohol moieties are often found in natural products and potent drugs, as seen in deoxyamino sugars, and they have also been used as synthetic intermediates. Thus, an effective method for the introduction of functionality to acyclic olefinic systems is required. While

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a variety of stereoselective synthetic methods especially for 1,2-amino alcohol moieties have been developed,⁷ there are relatively few methods for 1,3-amino alcohols.⁸ During our course of developing a simple synthetic strategy to deoxy-amino sugars, an intramolecular conjugate addition of γ -tri-chloroacetimidoyloxy- α , β -unsaturated esters was found.^{6,9} We here disclose the details of the trichloroacetimidate-mediated functionalization useful for the introduction of a nitrogen functionality¹⁰ on the β -carbon of γ - and δ -hydroxy- α , β -unsaturated esters, a new way to construct 1,2-amino or 1,3-amino alcohol moieties in an acyclic system.

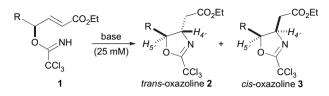
2. Results and discussion

2.1. Intramolecular conjugate additions of γ -trichloroacetimidoyloxy- α , β -unsaturated esters

In order to estimate this intramolecular conjugate addition, we first investigated the cyclization reaction of several kinds of γ -trichloroacetimidoyloxy- α , β -unsaturated esters (Table 1), which were prepared from the corresponding allylic alcohols¹¹ by treatment with 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) and trichloroacetonitrile in good to excellent yields.¹² When the substrate trichloroacetimidates **1a**–**d** were treated with

Table 1

Intramolecular conjugate addition of γ -trichloroacetimidoyloxy- α , β -unsaturated esters 1^a



Entry	Substrate R		Conditions	Product	
				Ratio 2/3	Yield ^b (%)
1	1a	Me	DBU (0.1 equiv), −10 °C	73:27 [°]	93
2	1a	Me	tert-BuOK (1.0 equiv), 0 °C	66:34 ^e	62
3	1a	Me	tert-BuOK (0.5 equiv), -78 °C	65:35 [°]	85
4	1a	Me	NaH (1.4 equiv), -20 °C	68:32 ^c	81
5	1b	TBSO	DBU (0.3 equiv), 0 °C	78:22 ^d	86
6	1b	TBSO	tert-BuOK (1.0 equiv), 0 °C	49:51 ^d	65
7	1b	TBSO	tert-BuOK (0.1 equiv), -20 °C	47:53 ^d	79
8	1b	TBSO	tert-BuOK (0.1 equiv), -78 °C	28:72 ^d	81
9 ^e	1c	TBSO	DBU (0.1 equiv), 0 °C	96:4 ^d	97
10 ^e	1c	TBSO	tert-BuOK (0.1 equiv), -62 °C	40:60 ^f	90
11	1d	HN O	DBU (1.6 equiv), -20 °C	86:14 ^c	66
12	1d	HN O	<i>tert</i> -BuOK (0.6 equiv), -100 °C	50:50°	82
13	1d	HN O	NaH (2.2 equiv), -20 °C	67:33 ^c	82

 $^{\rm a}$ All reactions were performed in CH₃CN (DBU) or THF (tert-BuOK, NaH).

^b Combined yield.

^d Products ratio calculated from their own isolated yield.

^e Experimental results obtained when *ent*-1c was used.

^f Separation of the isomers was not fully carried out and its ratio was determined by 270 MHz ¹H NMR spectroscopy.

a catalytic amount of DBU in CH₃CN, a facile cyclization occurred producing the oxazolines 2 (*trans*) and 3 (*cis*) (Table 1). The cis-trans stereochemistry of the oxazolines was determined according to the value of their proton coupling

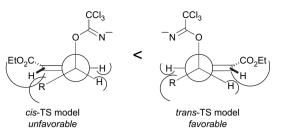


Figure 1. Proposed transition state models for cyclization.

constants $(J_{H-4'}, H-5')$,¹³ and it was also confirmed by nuclear Overhauser enhancement (NOE) experiments.⁶ In the case of substrate 1a, the ratio of 2a/3a was not affected by changing the conditions with maintained moderate trans-selectivity (entries 1-4). In the series with more sterically bulky R groups, when DBU was used as a base, higher trans-selectivity was observed (entry 5, 9, 11). Cyclization was also performed with stoichiometric amount of sodium hydride (NaH) in THF (entry 4, 13), showing little difference in the stereoselectivity. The trans-selectivity observed in the cyclization can be anticipated using the transition state (TS) models shown in Figure 1. The cyclization is thought to proceed through trans-TS model to give the *trans*-oxazolines, since cis-TS model is unfavorable than the trans-TS model due to its steric hindrance between the side chain R and the alkenyl group. With the exception of the compound 1c, trans-selectivity was not so high like the case of the carbamate-mediated intramolecular conjugate additions.¹⁴

Interestingly, reversal of diastereofacial selectivity was found (entry 6 and 10). In fact, upon treatment with potassium *tert*-butoxide (*tert*-BuOK) in THF, *cis*-oxazoline was dominantly produced. A notable temperature effect; namely, the ratio of *cis*-oxazoline increased with decreasing the reaction temperature, was also found (entries 6-8). These results indicated that *trans*-oxazoline was the thermodynamically controlled product as compared with the result of DBU. The reversal of diastereofacial selectivity was realized only at very low temperature as not reported in the carbamate-mediated intramolecular conjugate additions. It might be anticipated that the energy difference between trans-TS model and cis-TS model was somewhat smaller than those of the carbamate derivatives.

It is interesting to note that bis-trichloroacetimidate **1d** showed a marked tendency to cyclize five-membered oxazoline ring in the competitive cyclization between allylic and homoallylic trichloroacetimidates (entries 11-13). In this case, moderate trans-selectivity was observed by the treatment of DBU or NaH, while reversal of the selectivity was not fully realized with *tert*-BuOK even at -100 °C.

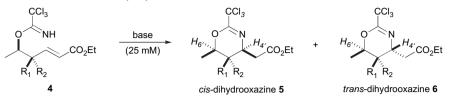
2.2. Intramolecular conjugate additions of δ -trichloroacetimidoyloxy- α , β -unsaturated esters

Next, we investigated the intramolecular conjugate additions of δ -trichloroacetimidoyloxy- α , β -unsaturated esters **4** (Table 2), which were prepared from the corresponding homoallylic alcohols.¹⁵ They were smoothly cyclized to six-membered cyclic compounds; namely, dihydrooxazines **5** (*cis*) and **6** (*trans*) by the treatment with *tert*-BuOK or NaH.

^c Inseparable products ratio determined by 270 MHz ¹H NMR spectroscopy.

Table 2

Intramolecular conjugate addition of δ -trichloroacetimidoyloxy- α , β -unsaturated esters 4^a



Entry	Substrate			Conditions	Product	
		R ₁	R ₂		Ratio 5/6	Yield ^b (%)
1	4 a	Н	Н	tert-BuOK (0.4 equiv), 0 °C	88:12 ^c	86
2	4a	Н	Н	NaH (1.1 equiv), 0 °C	87:13 [°]	86
3	4a	Н	Н	DBU (0.4 equiv), rt	_	d
4	4b ^e	Н	OMe	<i>tert</i> -BuOK (0.34 equiv), -20 °C	Exclusively cis	79
5	4b ^e	Н	OMe	NaH (1.2 equiv), −20 °C	Exclusively cis	88
6	4b ^e	Н	OMe	DBU (0.4 equiv), rt		d
7	4c	OPMP	Н	tert-BuOK (0.4 equiv), -20 °C	28:72 ^f	79
8	4c	OPMP	Н	NaH (1.1 equiv), 0 °C	24:76 ^f	89
9	4d	OTBS	Н	tert-BuOK (0.4 equiv), 0 °C	41:59 ^f	76
10	4d	OTBS	Н	NaH (1.1 equiv), 0 °C	28:72 ^f	79
11	4d	OTBS	Н	DBU (2.3 equiv), rt	—	g

^a All reactions were performed in CH₃CN (DBU) or THF (tert-BuOK, NaH).

^b Combined yield.

^c Inseparable products ratio determined by 270 MHz ¹H NMR spectroscopy.

^d Removal of *O*-trichloroacetimino group was mainly observed.

e Racemic material was used.

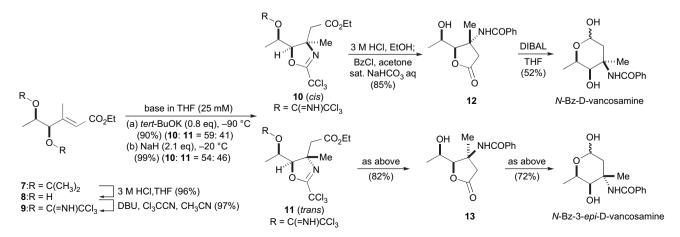
^f Products ratio calculated from their own isolated yield.

^g Removal of O-trichloroacetimino group and subsequent TBS group migration was observed.

It should be noted that in the δ -trichloroacetimidoyloxy series the cyclization was not realized with DBU even at rt, and gradual removal of *O*-trichloroacetimino group was mainly observed instead (entry 3, 6, 11). The stereochemistry of the obtained dihydrooxazines was determined by NOE experiments, and it was also confirmed according to the value of their proton coupling constants.¹⁶ In the case of substrate **4a**, cyclization was observed with moderate cis-selectivity (entry 1, 2). The γ -substituents played an important role for switching the diastereofacial selectivity. Actually, when a methoxy group was introduced at R₂ position, the cyclization proceeded with complete cis-selectivity (entry 4, 5), while when R₁ group was substituted by *p*-methoxyphenyloxy (PMPO) group or *tert*-butyldimethylsilyloxy (TBSO) group, the reaction was found to be carried out with moderate trans-selectivity (entries 7-10). Similar reversal of selectivity was also observed in the case of the carbamate-mediated intramolecular conjugate addition, and it was explained as cooperation of the steric and stereoelectronic effects.¹⁴

2.3. Concise synthesis of *D*-vancosamine and its 3-epimer

We planned to utilize the oxazoline ring formation for the synthesis of C-branched deoxyamino sugars, especially for vancosamine,¹⁷ the deoxyamino sugar constituent of vancomycin, and other antibiotics (Scheme 1). For the introduction



Scheme 1. Concise synthesis of D-vancosamine and 3-epi-D-vancosamine derivatives.

of the C(3) methyl-branched amino group of the vancosamine, we envisioned the methyl-branched bis-trichloroacetimidate 9 as a substrate for the intramolecular conjugate addition. The bis-imidate 9 was synthesized from the chiral diol 8, which was prepared from the known acetonide 7,¹⁸ in 97% yield. Bis-trichloroacetimidate 9 underwent a smooth conjugate addition constructing the oxazolines 10 (cis) and 11 (trans) by the treatment of tert-BuOK (10/11=59:41; 90% yield) or NaH (10/11=54:46; 99% yield) in THF. When DBU was used as a base, effective cyclization was not realized, since unknown by-products formation was observed while the starting imidate 9 still remained. The cis-trans stereochemistry of the oxazolines was confirmed by nuclear Overhauser enhancement (NOE) experiments,¹⁹ and it was ultimately determined by conversion to the vancosamine derivatives. Introduction of the methyl group on the double bond was expected to reduce the energy difference between trans-TS model and cis-TS model in Figure 1, but the effect was not so strong as expected. Then high cis-selectivity was not fully realized, however, the obtained oxazolines in high yield were fortunately separable, different from the case of 2d and 3d. Next, the hydrolysis of oxazoline ring and spontaneous γ -lactone formation and N-benzoylation were, respectively, carried out to give the γ -lactone 12 and 13 in good yields (85% for 12; 82% for 13). Final half reduction was successfully effected by diisopropylaluminum hydride (DIBAL) to produce N-Bz-D-vancosamine (52%) and N-Bz-3-epi-D-vancosamine (72%), respectively. The ratios of α - and β -anomers after ca. 1 month in DMSO- d_6 (α/β =ca. 1:3 for N-Bz-D-vancosamine; ca. 1:1 for N-Bz-3-epi-D-vancosamine) together with the detection of a small amount of the furanose forms were well accorded with that reported in the literature.²⁰ Additionally, their absolute values of specific optical rotation were also in good agreement with the enantiomers.²⁰

3. Conclusion

In conclusion, we have succeeded in developing a trichloroacetimidate-mediated method for the introduction of a nitrogen functionality on the β -carbon of γ - and δ -hydroxy- α , β -unsaturated esters, a novel way for constructing 1,2-amino or 1,3amino alcohol moieties in an acyclic system. Very concise synthesis of D-vancosamine derivatives and its 3-epimer was also achieved utilizing this methodology. Further investigations concerning the scope of this methodology are in progress.

4. Experimental section

4.1. General

All melting points are uncorrected. NMR spectra were recorded on a JEOL GSX-270 spectrometer. ¹H NMR and ¹³C NMR chemical shifts were reported in δ values based on internal tetramethylsilane ($\delta_{\rm H}$ =0) or solvent signal (CDCl₃ $\delta_{\rm C}$ =77.0; DMSO- $d_6 \delta_{\rm C}$ =39.5; C₆D₆ $\delta_{\rm H}$ =7.15) as reference. IR spectra were recorded on a HORIBA FT-720 Fouriertransform infrared spectrometer. Optical rotations were measured on a Rudolph Research Analytical AUTOPOL V polarimeter and $[\alpha]_D$ values are given in units of $10^{-1} \text{ deg cm}^2 \text{ g}^{-1}$. Mass spectra were measured on a Applied Biosystems QSTAR XL hybrid Q-TOF or a JEOL JMS-SX102A (FAB and EI) mass spectrometer. Flash silica gel column chromatography was carried out on KANTO CHEM-ICAL CO., INC. Silica Gel 60 N (spherical, neutral, 40– 50 µm) or Merck Kieselgel 60 (230–400 mesh), Art. Nr. 9385.

4.2. General procedure for the trichloroacetimidates

To a cooled (bath temp below -20 °C) solution of corresponding alcohol and trichloroacetonitrile (10 equiv) in acetonitrile was added dropwise DBU (ca. 1.1–1.2 equiv), and the mixture was stirred for ca. 30 min under a dry atmosphere (calcium chloride tube). The reaction mixture was poured into cold saturated aq NH₄Cl and extracted with EtOAc. The extract was washed successively with saturated aq NH₄Cl and brine, dried (MgSO₄), and concentrated in vacuo. The residue was purified by flash column chromatography to give the trichloroacetimidate.

4.3. Data of the trichloroacetimidates

4.3.1. Ethyl (2E,4S)-4-trichloroacetimidoyloxypent-2enoate (**1***a*)

Yield: 77%; colorless oil; $[\alpha]_D^{27.1}$ -0.637 (*c* 1.57, CHCl₃); ν_{max} (neat) 3344, 2983, 1722, 1664, 1308, 1282, 1182, 1076, 1043, 976, 796, 648 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 8.39 (br s, 1H), 6.96 (dd, *J*=4.5, 15.8 Hz, 1H), 6.09 (dd, *J*=1.7, 15.6 Hz, 1H), 5.65 (ddq, *J*=1.7, 4.7, 6.6 Hz, 1H), 4.21 (q, *J*=7.1 Hz, 2H), 1.49 (d, *J*=6.6 Hz, 3H), 1.30 (t, *J*=7.2 Hz, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 166.1, 161.5, 145.5, 121.1, 91.3, 73.6, 60.6, 19.0, 14.2. Anal. Calcd for C₉H₁₂Cl₃NO₃: C, 37.46; H, 4.19; N, 4.85. Found: C, 37.02; H, 3.89; N, 4.66%.

4.3.2. Ethyl (2E,4R,5R)-5-tert-butyldimethylsilyloxy-4trichloroacetimidoyloxyhex-2-enoate (**1b**)

Yield: 99%; colorless oil; $[\alpha]_D^{24.5} + 40.0$ (*c* 1.17, CHCl₃); ν_{max} (neat) 3348, 2956, 2931, 2858, 1724, 1670, 1660, 1311, 1259, 1180, 1099, 1078, 1049, 835, 798, 777, 648 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 8.43 (br s, 1H), 7.05 (dd, *J*=4.0, 15.9 Hz, 1H), 6.14 (dd, *J*=1.8, 15.9 Hz, 1H), 5.52 (ddd, *J*=1.9, 4.6, 4.6 Hz, 1H), 4.23–4.10 (m, 1H), 4.21 (q, *J*=7.0 Hz, 2H), 1.30 (t, *J*=7.0 Hz, 3H), 1.15 (d, *J*=6.2 Hz, 3H), 0.90 (s, 9H), 0.11 (s, 3H), 0.10 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 165.9, 161.9, 141.8, 122.6, 91.2, 79.5, 67.5, 60.5, 25.7, 18.5, 17.9, 14.2, -4.9. Anal. Calcd for C₁₆H₂₈Cl₃NO₄Si: C, 44.40; H, 6.52; N, 3.24. Found: C, 44.47; H, 6.23; N, 3.15%.

4.3.3. Ethyl (2E,4S,5R)-5-tert-butyldimethylsilyloxy-4-

trichloroacetimidoyloxyhex-2-enoate (ent-1c)

Yield: 94%; colorless oil; $[\alpha]_D^{23.7}$ –7.60 (*c* 2.31, CHCl₃); ν_{max} (neat) 3348, 2956, 2931, 2858, 1724, 1670, 1377, 1311, 1259, 1180, 1117, 1080, 1047, 984, 831, 796, 777, 648 cm⁻¹; ¹H

NMR (270 MHz, CDCl₃) δ 8.40 (br s, 1H), 7.01 (dd, *J*=5.1, 16.0 Hz, 1H), 6.07 (dd, *J*=1.5, 15.8 Hz, 1H), 5.39 (ddd, *J*=1.5, 5.1, 5.1 Hz, 1H), 4.20 (q, *J*=7.1 Hz, 2H), 4.06 (dq, *J*=5.3, 6.3 Hz, 1H), 1.29 (t, *J*=7.2 Hz, 3H), 1.25 (d, *J*=6.4 Hz, 3H), 0.88 (s, 9H), 0.07 (s, 6H); ¹³C NMR (67.8 MHz, CDCl₃) δ 165.8, 161.7, 142.3, 122.9, 91.3, 81.0, 69.8, 60.5, 25.7, 20.1, 17.9, 14.2, -4.5, -5.0. Anal. Calcd for C₁₆H₂₈Cl₃NO₄Si: C, 44.40; H, 6.52; N, 3.24. Found: C, 44.70; H, 6.31; N, 3.12%.

4.3.4. Ethyl (2E,4R,5R)-4,5-bis(trichloroacetimidoyloxy)hex-2-enoate (1d)

Yield: 96%; colorless oil; $[\alpha]_D^{28.1} + 26.1$ (*c* 0.980, CHCl₃); ν_{max} (neat) 3344, 2985, 1724, 1664, 1300, 1282, 1184, 1074, 831, 796, 648, 490 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 8.52 (br s, 1H), 8.44 (br s, 1H), 6.98 (dd, *J*=4.7, 15.8 Hz, 1H), 6.21 (dd, *J*=1.7, 15.8 Hz, 1H), 5.86 (ddd, *J*=1.7, 4.9, 4.9 Hz, 1H), 5.41 (dq, *J*=5.3, 6.4 Hz, 1H), 4.21 (dq, *J*=7.2, 11.0 Hz, 1H), 4.20 (dq, *J*=7.1, 10.9 Hz, 1H), 1.42 (d, *J*=6.4 Hz, 3H), 1.29 (t, *J*=7.2 Hz, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 165.5, 161.6, 161.3, 140.0, 124.2, 91.2, 90.9, 76.7, 74.1, 60.7, 14.5, 14.2. Anal. Calcd for C₁₂H₁₄Cl₆N₂O₄: C, 31.13; H, 3.05; N, 6.05. Found: C, 31.12; H, 3.08; N, 5.89%.

4.3.5. *Ethyl* (2E,5R)-5-(*trichloroacetimidoyloxy*)*hex-2enoate* (**4***a*)

Yield: 85%; colorless oil; $[\alpha]_D^{29.4}$ +6.12 (*c* 1.05, CHCl₃); ν_{max} (neat) 3344, 2981, 1720, 1662, 1321, 1306, 1292, 1273, 1180, 1080, 1057, 1045, 980, 796, 648 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 8.31 (br s, 1H), 6.95 (ddd, *J*=7.4, 7.4, 15.6 Hz, 1H), 5.91 (ddd, *J*=1.4, 1.4, 15.7 Hz, 1H), 5.16 (ddq, *J*=6.2, 6.2, 6.2 Hz, 1H), 4.18 (q, *J*=7.1 Hz, 2H), 2.63 (dddd, *J*=1.4, 6.5, 7.4, 14.7 Hz, 1H), 2.59 (dddd, *J*=1.6, 5.6, 7.4, 14.7 Hz, 1H), 1.82 (d, *J*=6.2 Hz, 3H), 1.28 (t, *J*=7.2 Hz, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 166.1, 161.9, 143.2, 124.4, 91.6, 74.2, 60.3, 37.9, 18.8, 14.2. Anal. Calcd for C₁₀H₁₄Cl₃NO₃: C, 39.69; H, 4.66; N, 4.63. Found: C, 39.64; H, 4.38; N, 4.54%.

4.3.6. *Ethyl* (2*E*,4*S**,5*R**)-5-(*trichloroacetimidoyloxy*)-4*methoxyhex*-2-*enoate* (**4***b*)

Yield: 96%; colorless oil; ν_{max} (neat) 3344, 2985, 1722, 1664, 1300, 1176, 1078, 796, 648 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 8.34 (br s, 1H), 6.85 (dd, *J*=5.9, 15.7 Hz, 1H), 6.11 (dd, *J*=1.5, 15.8 Hz, 1H), 5.10 (dq, *J*=4.3, 6.4 Hz, 1H), 4.21 (q, *J*=7.1 Hz, 2H), 4.03 (ddd, *J*=1.5, 4.3, 5.9 Hz, 1H), 3.42 (s, 3H), 1.35 (d, *J*=6.6 Hz, 3H), 1.29 (t, *J*=7.2 Hz, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 165.8, 161.8, 143.8, 124.1, 91.5, 81.8, 76.6, 60.5, 58.3, 14.2, 14.1. Anal. Calcd for C₁₁H₁₆Cl₃NO₄: C, 39.72; H, 4.85; N, 4.21. Found: C, 39.41; H, 4.79; N, 3.99%.

4.3.7. *Ethyl* (2*E*,4*R*,5*R*)-5-(*trichloroacetimidoyloxy*)-4-(4-*methoxyphenoxy*)*hex-2-enoate* (4*c*)

Yield: 94%; colorless oil; $[\alpha]_D^{25.1}$ +17.1 (*c* 0.575, CHCl₃); ν_{max} (neat) 3340, 2985, 1720, 1664, 1508, 1304, 1227, 1180, 1059, 796, 648 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 8.43 (br s, 1H), 7.03 (dd, *J*=4.6, 15.7 Hz, 1H), 6.95–6.87 (m, 2H), 6.85–6.77 (m, 2H), 6.19 (dd, *J*=1.6, 15.7 Hz, 1H), 5.36 (dq, *J*=5.3, 6.4 Hz, 1H), 4.99 (ddd, *J*=1.6, 5.0, 5.0 Hz, 1H), 4.20 (dq, *J*=7.1, 11.2 Hz, 1H), 4.19 (dq, *J*=7.1, 11.4 Hz, 1H), 3.76 (s, 3H), 1.40 (d, *J*=6.4 Hz, 3H), 1.28 (t, *J*=7.2 Hz, 3H); 13 C NMR (67.8 MHz, CDCl₃) δ 165.8, 161.9, 154.5, 151.7, 142.4, 124.3, 117.1, 114.6, 91.3, 77.9, 75.2, 60.6, 55.6, 14.3, 14.2. Anal. Calcd for C₁₇H₂₀Cl₃NO₅: C, 48.08; H, 4.75; N, 3.30. Found: C, 47.99; H, 4.63; N, 3.18%.

4.3.8. Ethyl (2E,4R,5R)-4-tert-butyldimethylsilyloxy-5-(trichloroacetimidoyloxy)hex-2-enoate (4d)

Yield: 96%; colorless oil; $[\alpha]_{25.9}^{25.9}$ +55.7 (*c* 0.620, CHCl₃); ν_{max} (neat) 3348, 2956, 2931, 1724, 1664, 1298, 1288, 1261, 1084, 1063, 835, 798, 650 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 8.38 (br s, 1H), 7.02 (dd, *J*=4.1, 15.6 Hz, 1H), 6.11 (dd, *J*=1.9, 15.6 Hz, 1H), 5.07 (dq, *J*=5.0, 6.4 Hz, 1H), 4.65 (ddd, *J*=1.8, 4.1, 4.8 Hz, 1H), 4.21 (dd, *J*=7.1, 10.9 Hz, 1H), 4.20 (dq, *J*=7.1, 10.9 Hz, 1H), 1.30 (t, *J*=7.2 Hz, 3H), 1.25 (d, *J*=6.4 Hz, 3H), 0.93 (s, 9H), 0.13 (s, 3H), 0.08 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 166.3, 161.9, 146.1, 122.6, 91.5, 76.9, 70.8, 60.4, 25.7, 18.1, 14.2, 13.3, -4.9, -5.0. Anal. Calcd for C₁₆H₂₈Cl₃NO₄Si: C, 44.40; H, 6.52; N, 3.24. Found: C, 44.56; H, 6.48; N, 3.11%.

4.4. Typical procedure of the conjugate additions using DBU: ethyl 2-((4R,5S)-2-(trichloromethyl)-4,5-dihydro-5-((1R)-tertbutyldimethylsilyloxyethyl)oxazol-4-yl)acetate (ent-2c) and ethyl 2-((4S,5S)-2-(trichloromethyl)-4,5-dihydro-5-((1R)-tertbutyldimethylsilyloxyethyl)oxazol-4-yl)acetate (ent-3c)

To an ice-cooled solution of the trichloroacetimidate *ent*-1c (58.2 mg, 0.134 mmol) in acetonitrile (5.5 mL) was added dropwise DBU (2.0 μ L, 0.013 mmol), and the mixture was stirred for ca. 16 h under Ar atmosphere. The reaction mixture was poured into cold saturated aq NH₄Cl and extracted with EtOAc. The extract was washed with brine, dried (MgSO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (hexane/EtOAc=5:1) to give *trans*-oxazoline *ent*-2c (colorless oil, less polar, 53.4 mg, 92%) and *cis*-oxazoline *ent*-3c (colorless oil, more polar, 2.0 mg, 3.4%).

4.4.1. trans-Oxazoline ent-2c

[α]_D^{27.4} +40.8 (*c* 0.495, CHCl₃); ν_{max} (neat) 2956, 2931, 2858, 1736, 1664, 1375, 1257, 1182, 1159, 1115, 1078, 1032, 991, 837, 795, 777 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 4.57 (ddd, *J*=5.6, 5.8, 6.6 Hz, 1H), 4.51 (dd, *J*=4.5, 6.6 Hz, 1H), 4.165 (dq, *J*=7.2, 10.8 Hz, 1H), 4.164 (dq, *J*=7.1, 10.8 Hz, 1H), 4.02 (dq, *J*=4.4, 6.3 Hz, 1H), 2.72 (dd, *J*=5.8, 15.6 Hz, 1H), 2.66 (dd, *J*=5.6, 15.8 Hz, 1H), 1.27 (t, *J*=7.2 Hz, 3H), 1.20 (d, *J*=6.4 Hz, 3H), 0.88 (s, 9H), 0.10 (s, 3H), 0.09 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 170.1, 162.4, 91.0, 86.5, 68.6, 64.6, 60.8, 39.3, 25.8, 19.3, 17.9, 14.2, -4.1, -4.8. Anal. Calcd for C₁₆H₂₈Cl₃NO₄Si: C, 44.40; H, 6.52; N, 3.24. Found: C, 44.67; H, 6.37; N, 3.08%.

4.4.2. cis-Oxazoline ent-3c

 $[\alpha]_D^{27.5}$ -40.1 (*c* 0.635, CHCl₃); ν_{max} (neat) 2956, 2931, 2858, 1736, 1662, 1379, 1257, 1180, 1107, 1036, 993, 835, 793, 777, 669 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 4.83

(ddd, J=5.8, 8.1, 9.6 Hz, 1H), 4.68 (dd, J=5.3, 9.6 Hz, 1H), 4.194 (dq, J=7.2, 10.9 Hz, 1H), 4.190 (dq, J=5.6, 6.4 Hz, 1H), 4.18 (dq, J=7.1, 10.8 Hz, 1H), 3.00 (dd, J=5.9, 16.3 Hz, 1H), 2.65 (dd, J=8.0, 16.3 Hz, 1H), 1.30 (d, J=6.4 Hz, 3H), 1.21 (t, J=7.1 Hz, 3H), 0.89 (s, 9H), 0.108 (s, 3H), 0.105 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 171.1, 162.3, 91.0, 88.7, 67.1, 64.5, 60.8, 35.1, 25.8, 20.5, 17.9, 14.2, -3.8, -4.7. Anal. Calcd for C₁₆H₂₈Cl₃NO₄Si: C, 44.40; H, 6.52; N, 3.24. Found: C, 44.63; H, 6.55; N, 3.09%.

4.5. Typical procedure of the conjugate additions using tert-BuOK: ethyl 2-((4S,5R)-2-(trichloromethyl)-4,5-dihydro-5-((1R)-tert-butyldimethylsilyloxyethyl)oxazol-4-yl)acetate (**2b**) and ethyl 2-((4R,5R)-2-(trichloromethyl)-4,5-dihydro-5-((1R)-tert-butyldimethylsilyloxyethyl)oxazol-4-yl)acetate (**3b**)

To a solution of the trichloroacetimidate **1b** (56.0 mg, 0.129 mmol) in THF (5.5 mL) was added *tert*-BuOK (1.7 mg, 0.015 mmol) in one portion under Ar atmosphere at -78 °C, and the mixture was stirred for ca. 2 h. The reaction mixture was poured into cold saturated aq NH₄Cl and extracted with EtOAc. The extract was washed with brine, dried (MgSO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (hexane/EtOAc=11:1) to give *trans*-oxazoline **2b** (colorless solid, less polar, 12.9 mg, 23%) and *cis*-oxazoline **3b** (colorless oil, more polar, 32.7 mg, 58%). Analytical sample (colorless powder) of **2b** was obtained by recrystallization from MeOH (-15 °C).

4.5.1. trans-Oxazoline 2b

Mp 52.0–52.5 °C; $[α]_D^{27.6}$ –78 (*c* 0.18, CHCl₃); $ν_{max}$ (KBr) 2958, 2931, 2858, 1728, 1664, 1375, 1244, 1186, 1149, 1063, 1038, 941, 920, 839, 798, 777, 663 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 4.51 (dd, *J*=2.6, 6.0 Hz, 1H), 4.48 (ddd, *J*=4.3, 6.0, 8.1 Hz, 1H), 4.17 (q, *J*=7.1 Hz, 2H), 4.05 (dq, *J*=2.4, 6.4 Hz, 1H), 2.84 (dd, *J*=4.3, 16.7 Hz, 1H), 2.60 (dd, *J*=8.4, 16.6 Hz, 1H), 1.28 (t, *J*=7.2 Hz, 3H), 1.26 (d, *J*=6.4 Hz, 3H), 0.87 (s, 9H), 0.10 (s, 3H), 0.08 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 170.5, 162.5, 91.3, 86.6, 68.8, 64.8, 60.8, 39.0, 25.6, 19.0, 17.9, 14.2, -4.2, -5.0. Anal. Calcd for C₁₆H₂₈Cl₃NO₄Si: C, 44.40; H, 6.52; N, 3.24. Found: C, 44.47; H, 6.31; N, 3.18%.

4.5.2. cis-Oxazoline 3b

[α]_D^{27.3} -41.7 (*c* 0.620, CHCl₃); ν_{max} (neat) 2958, 2931, 2858, 1732, 1664, 1329, 1255, 1184, 1093, 1030, 1001, 926, 837, 793, 777, 671 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 4.80 (dd, *J*=2.1, 9.6 Hz, 1H), 4.72 (ddd, *J*=5.3, 9.2, 9.6 Hz, 1H), 4.20 (dq, *J*=7.2, 10.7 Hz, 1H), 4.17 (dq, *J*=7.2, 10.7 Hz, 1H), 4.01 (dq, *J*=2.1, 6.4 Hz, 1H), 3.13 (dd, *J*=9.2, 17.3 Hz, 1H), 3.03 (dd, *J*=5.3, 17.3 Hz, 1H), 1.31 (d, *J*=6.4 Hz, 3H), 1.28 (t, *J*=7.2 Hz, 3H), 0.89 (s, 9H), 0.09 (s, 3H), 0.08 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 171.7, 162.4, 88.9, 86.8, 67.6, 64.8, 60.9, 34.3, 26.0, 20.6, 18.0, 14.2, -3.1, -4.9. Anal. Calcd for C₁₆H₂₈Cl₃NO₄Si: C, 44.40; H, 6.52; N, 3.24. Found: C, 44.77; H, 6.33; N, 2.99%.

4.6. Data of the oxazolines

4.6.1. Ethyl 2-((4S,5S)-2-(trichloromethyl)-4,5-dihydro-5-methyloxazol-4-yl)acetate (**2a**) and ethyl 2-((4R,5S)-2-(trichloromethyl)-4,5-dihydro-5-methyloxazol-4-yl)acetate (**3a**)

Colorless oil; $[\alpha]_{D}^{28.6} - 35.6$ (*c* 1.00, CHCl₃, sample cyclized with DBU); ν_{max} (neat) 2983, 1734, 1658, 1375, 1238, 1184, 1024, 893, 829, 795, 665 cm⁻¹; ¹H NMR (270 MHz, C₆D₆) δ major (trans) 4.26 (dq, *J*=6.3, 6.3 Hz, 1H), 3.94 (ddd, *J*=5.0, 6.3, 9.1 Hz, 1H), 3.81 (q, *J*=7.3 Hz, 2H), 2.46 (dd, *J*=4.9, 16.7 Hz, 1H), 2.03 (dd, *J*=9.1, 16.6 Hz, 1H), 1.04 (d, *J*= 6.2 Hz, 3H), 0.863 (t, *J*=7.1 Hz, 3H); minor (cis) 4.55 (dq, *J*=6.4, 9.2 Hz, 1H), 4.43 (ddd, *J*=6.3, 8.7, 9.2 Hz, 1H), 3.88– 3.73 (overlapping m, 2H), 2.50 (dd, *J*=6.3, 16.8 Hz, 1H), 2.14 (dd, *J*=8.5, 16.7 Hz, 1H), 0.865 (t, *J*=7.2 Hz, 3H), 0.77 (d, *J*=6.6 Hz, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ major (trans) 170.4, 162.2, 86.6, 85.6, 69.5, 60.9, 38.7, 20.4, 14.14; minor (cis) 170.9, 162.3, 86.7, 82.9, 65.0, 61.0, 34.5, 14.5, 14.11. Anal. Calcd for C₉H₁₂Cl₃NO₃: C, 37.46; H, 4.19; N, 4.85. Found: C, 37.38; H, 4.23; N, 4.58%.

4.6.2. Ethyl 2-((4S,5R)-5-(R)-1-(trichloroacetimidoyloxy)ethyl)-2-(trichloromethyl)-4,5-dihydro-oxazol-4yl)acetate (**2d**) and ethyl 2-((4R,5R)-5-(R)-1-(trichloroacetimidoyloxy)ethyl)-2-(trichloromethyl)-4,5-dihydrooxazol-4-yl)acetate (**3d**)

Colorless oil; $[\alpha]_D^{28.2}$ –53 (c 0.42, CHCl₃, sample cyclized with *tert*-BuOK at -100 °C); ν_{max} (neat) 3344, 2983, 1732, 1668, 1375, 1348, 1300, 1284, 1236, 1186, 1080, 1032, 987, 930, 839, 795, 650 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ major (trans) 8.43 (s, 1H), 5.35 (dq, J=2.2, 6.5 Hz, 1H), 4.78 (dd, J=2.2, 5.7 Hz, 1H), 4.50 (ddd, J=4.1, 5.8, 9.8 Hz, 1H), 4.18 (q, J=7.2 Hz, 2H), 2.92 (dd, J=4.0, 17.0 Hz, 1H), 2.62 (dd, J=9.8, 16.9 Hz, 1H), 1.50 (d, J=6.4 Hz, 3H), 1.28 (t, J=7.2 Hz, 3H); minor (cis) 8.49 (s, 1H), 5.18 (br q, J=6.6 Hz, 1H), 5.12 (dd, J=1.0, 9.9 Hz, 1H), 4.82 (ddd, J=4.9, 9.9, 10.7 Hz, 1H), 4.11-4.28 (overlapping, 2H), 3.11 (dd, J=5.0, 17.8 Hz, 1H), 2.71 (dd, J=10.7, 18.0 Hz, 1H), 1.46 (d, J=6.4 Hz, 3H), 1.28 (t, J=7.2 Hz, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ major (trans) 170.4, 162.3, 160.8, 91.1, 89.2, 86.3, 75.2, 65.3, 61.0, 38.5, 14.7, 14.1; minor (cis) 171.5, 162.0, 161.8, 90.9, 86.7, 86.4, 73.1, 64.9, 61.1, 34.2, 15.2, 14.1; HRMS (ESI-TOF) calcd for $C_{12}H_{15}Cl_6N_2O_4$ [M+H]⁺: 460.9157, found: 460.9157.

4.7. Data of the dihydrooxazines

4.7.1. Ethyl 2-((4R,6R)-2-(trichloromethyl)-5,6-dihydro-6methyl-4H-1,3-oxazin-4-yl)acetate (5a) and ethyl 2-((4S,6R)-2-(trichloromethyl)-5,6-dihydro-6-methyl-4H-1,3-oxazin-4-yl)acetate (6a)

Colorless oil; $[\alpha]_D^{29.3}$ +26.0 (*c* 1.11, CHCl₃, sample cyclized with *tert*-BuOK); ν_{max} (neat) 2981, 1734, 1676, 1446, 1375, 1340, 1240, 1203, 1180, 1122, 1026, 958, 791, 667 cm⁻¹; ¹H NMR (270 MHz, C₆D₆) δ major (cis) 3.96 (q, *J*=7.1 Hz, 2H), 3.72 (dddd, *J*=4.6, 6.5, 7.9, 11.1 Hz, 1H), 3.57 (ddq, *J*=2.6,

6.3, 11.4 Hz, 1H), 2.56 (dd, J=6.3, 15.7 Hz, 1H), 2.13 (dd, J=7.8, 15.7 Hz, 1H), 1.36 (ddd, J=2.6, 4.5, 13.5 Hz, 1H), 0.96 (t, J=7.2 Hz, 3H), 0.84 (d, J=6.2 Hz, 3H), 0.73 (ddd, J=11.3, 11.3, 13.6 Hz, 1H); minor (trans) 4.02–3.80 (overlapping m, 4H), 2.52 (dd, J=6.2, 15.4 Hz, 1H), 2.06 (dd, J=8.3, 15.4 Hz, 1H), 1.17 (ddd, J=4.6, 6.5, 14.0 Hz, 1H), 1.15 (ddd, J=5.7, 5.7, 14.0 Hz, 1H), 0.94 (t, J=7.2 Hz, 3H), 0.80 (d, J=6.4 Hz, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ major (cis) 171.3, 154.1, 92.3, 74.3, 60.56, 51.1, 41.4, 34.4, 20.8, 14.2; minor (trans) 171.0, 153.8, 92.5, 71.2, 60.65, 47.6, 40.8, 31.4, 20.2, 14.2. Anal. Calcd for C₁₀H₁₄Cl₃NO₃: C, 39.69; H, 4.66; N, 4.63. Found: C, 40.02; H, 4.59; N, 4.48%.

4.7.2. *Ethyl* 2-((4S*,5S*,6R*)-2-(trichloromethyl)-5,6dihydro-5-methoxy-6-methyl-4H-1,3-oxazin-4-yl)acetate (**5b**)

Colorless oil; ν_{max} (neat) 2983, 1736, 1684, 1215, 1124, 1099, 818, 793, 665 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 4.18 (q, *J*=7.1 Hz, 2H), 4.14 (dq, *J*=6.3, 9.2 Hz, 1H), 3.82 (ddd, *J*=5.8, 6.5, 8.9 Hz, 1H), 3.55 (s, 3H), 3.11 (dd, *J*=9.0, 9.0 Hz, 1H), 2.79 (dd, *J*=5.8, 14.7 Hz, 1H), 2.64 (dd, *J*=6.5, 14.9 Hz, 1H), 1.53 (d, *J*=6.2 Hz, 3H), 1.27 (t, *J*=7.1 Hz, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 171.0, 152.7, 91.6, 79.4, 76.5, 60.6, 60.5, 56.6, 38.1, 17.8, 14.2. Anal. Calcd for C₁₁H₁₆Cl₃NO₄: C, 39.72; H, 4.85; N, 4.21. Found: C, 39.85; H, 4.85; N, 4.13%.

4.7.3. *Ethyl* 2-((4*S*,5*R*,6*R*)-2-(*trichloromethyl*)-5,6-*dihydro*-5-(4-*methoxyphenoxy*)-6-*methyl*-4*H*-1,3-*oxazin*-4-*yl*)*acetate* (5*c*)

Colorless oil; $[\alpha]_D^{27.6}$ +5.0 (*c* 0.10, CHCl₃); ν_{max} (neat) 2933, 1728, 1682, 1506, 1225, 1180, 1039, 793 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 6.89–6.76 (m, 4H), 4.70 (dd, *J*=1.3, 3.2 Hz, 1H), 4.60 (ddq, *J*=1.3, 1.3, 6.6 Hz, 1H), 4.21 (dddd, *J*=1.3, 3.0, 5.3, 9.8 Hz, 1H), 4.05 (dq, *J*=7.2, 10.7 Hz, 1H), 4.02 (dq, *J*=7.1, 10.7 Hz, 1H), 3.76 (s, 3H), 2.83 (dd, *J*=5.3, 17.1 Hz, 1H), 2.74 (dd, *J*=9.8, 17.3 Hz, 1H), 1.42 (d, *J*=6.4 Hz, 3H), 1.14 (t, *J*=7.2 Hz, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 171.8, 154.3, 153.6, 153.3, 116.5, 114.7, 91.9, 76.3, 71.0, 60.6, 55.7, 55.2, 36.5, 17.1, 14.1; HRMS (EI) calcd for C₁₇H₂₀Cl₃NO₅ [M]⁺: 423.04071, found: 423.0405.

4.7.4. Ethyl 2-((4*R*,5*R*,6*R*)-2-(*trichloromethyl*)-5,6-*dihydro*-5-(4-*methoxyphenoxy*)-6-*methyl*-4*H*-1,3-*oxazin*-4-*yl*)*acetate* (**6***c*)

Colorless oil; $[\alpha]_{D}^{27.2}$ +36.7 (*c* 0.570, CHCl₃); ν_{max} (neat) 2983, 1732, 1682, 1506, 1223, 1184, 1036, 823, 793, 671 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 6.98–6.90 (m, 2H), 6. 87–6.80 (m, 2H), 4.63 (dq, *J*=3.1, 6.5 Hz, 1H), 4.39 (dd, *J*=3.1, 4.8 Hz, 1H), 4.22 (ddd, *J*=4.8, 5.5, 8.3 Hz, 1H), 4.17 (dq, *J*=7.2, 10.8 Hz, 1H), 4.16 (dq, *J*=7.1, 10.8 Hz, 1H), 3.77 (s, 3H), 2.78 (dd, *J*=5.6, 15.6 Hz, 1H), 2.57 (dd, *J*=8.3, 15.6 Hz, 1H), 1.48 (d, *J*=6.4 Hz, 3H), 1.24 (t, *J*=7.1 Hz, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 170.5, 154.8, 153.2, 150.7, 117.6, 114.8, 91.9, 72.7, 71.6, 60.9, 55.7, 52.1, 38.4, 15.1, 14.2; HRMS (EI) calcd for C₁₇H₂₀Cl₃NO₅ [M]⁺: 423.04071, found: 423.0407.

4.7.5. Ethyl 2-((4S,5R,6R)-5-tert-butyldimethylsilyloxy-2-(trichloromethyl)-5,6-dihydro-6-methyl-4H-1,3-oxazin-4-yl)acetate (5d)

Colorless oil; $[\alpha]_D^{26.6}$ -0.198 (*c* 0.505, CHCl₃); ν_{max} (neat) 2931, 1732, 1684, 1375, 1255, 1228, 1180, 1165, 1057, 839, 795 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 4.44 (ddq, *J*=1.2, 1.2, 6.6 Hz, 1H), 4.17 (q, *J*=7.1 Hz, 2H), 4.04 (dd, *J*=1.1, 2.4 Hz, 1H), 4.01 (dddd, *J*=1.3, 2.5, 5.8, 8.6 Hz, 1H), 2.74 (dd, *J*=5.8, 17.5 Hz, 1H), 2.70 (dd, *J*=8.7, 17.2 Hz, 1H), 1.39 (d, *J*=6.4 Hz, 3H), 1.28 (t, *J*=7.2 Hz, 3H), 0.90 (s, 9H), 0.10 (s, 3H), 0.03 (s, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 172.0, 153.2, 92.0, 77.04, 65.6, 60.5, 56.1, 36.8, 25.8, 18.3, 17.4, 14.2, -3.9, -4.2; HRMS (FAB) calcd for C₁₆H₂₉Cl₃NO₄Si [M+H]⁺: 432.09314, found: 432.0931.

4.7.6. Ethyl 2-((4R,5R,6R)-5-tert-butyldimethylsilyloxy-2-(trichloromethyl)-5,6-dihydro-6-methyl-4H-1,3-oxazin-4yl)acetate (**6d**)

Colorless oil; $[\alpha]_{D}^{25.5}$ +54.1 (*c* 0.765, CHCl₃); ν_{max} (neat) 2956, 2931, 1736, 1682, 1261, 1223, 1128, 839, 795, 667 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 4.38 (dq, *J*=2.9, 6.5 Hz, 1H), 4.182 (dq, *J*=7.2, 10.8 Hz, 1H), 4.178 (dq, *J*=7.0, 11.3 Hz, 1H), 3.95 (ddd, *J*=4.6, 5.6, 8.2 Hz, 1H), 3.83 (dd, *J*=2.9, 4.6 Hz, 1H), 2.68 (dd, *J*=5.8, 15.6 Hz, 1H), 2.48 (dd, *J*=8.2, 15.7 Hz, 1H), 1.37 (d, *J*=6.4 Hz, 3H), 1.28 (t, *J*=7.2 Hz, 3H), 0.89 (s, 9H), 0.14 (s, 3H), 0.10 (s, 3H); 1³C NMR (67.8 MHz, CDCl₃) δ 170.6, 153.0, 92.1, 73.1, 66.8, 60.8, 55.4, 38.3, 25.6, 17.9, 15.2, 14.2, -4.7, -4.8. Anal. Calcd for C₁₆H₂₈Cl₃NO₄Si: C, 44.40; H, 6.52; N, 3.24. Found: C, 44.70; H, 6.58; N, 3.09%.

4.8. Syntheses of N-Bz-3-D-vancosamine and N-Bz-3-epi-D-vancosamine

4.8.1. Ethyl (2*E*,4*R*,5*R*)-4,5-dihydroxy-3-methylhex-2enoate (**8**)

To a solution of the known acetonide 7 (1.35 g, 5.91 mmol) in THF (32 mL) was added dropwise 3 M HCl (32 mL), and the mixture was stirred for 2.5 h. Then excess amount of NaHCO₃ powder was carefully added portionwise to the reaction mixture. The aqueous mixture was saturated with NaCl, and extracted with EtOAc and CH₂Cl₂. The extract was dried (MgSO₄) and concentrated in vacuo. The residue was purified by flash column chromatography (hexane/EtOAc=1:2) to give diol 8 (1.07 g, 96%) as a colorless solid. Analytical sample (colorless powder) was obtained by recrystallization from EtOAc/hexane (0 °C); mp 40.0–43.5 °C; $[\alpha]_D^{29.4}$ –0.995 (c 1.01, CHCl₃); v_{max} (KBr) 3446, 3396, 2981, 1709, 1647, 1223, 1153, 1038, 860 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 5.95-5.91 (m, 1H), 4.17 (q, J=7.1 Hz, 2H), 3.91-3.79 (m, 2H), 2.81 (br d, 1H), 2.43 (br s, 1H), 2.13 (d, J=1.5 Hz, 3H), 1.29 (t, J=7.1 Hz, 3H), 1.20 (d, J=6.2 Hz, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 166.5, 156.9 117.6, 81.0, 68.4, 60.0, 19.4, 15.1, 14.2. Anal. Calcd for C₉H₁₆O₄: C, 57.43; H, 8.57. Found: C, 57.46; H, 8.61%.

4.8.2. *Ethyl* (2E,4R,5R)-4,5-bis(trichloroacetimidoyloxy)-3methylhex-2-enoate (9)

To a cooled (bath temp below -20 °C) solution of the diol 8 (122.2 mg, 0.649 mmol) and trichloroacetonitrile (1.30 mL, 13.0 mmol) in acetonitrile (3.5 mL) was added dropwise DBU (213.5 µL, 1.43 mmol), and the mixture was stirred for ca. 30 min under a dry atmosphere (calcium chloride tube). The reaction mixture was poured into cold saturated aq NH₄Cl and extracted with EtOAc. The extract was washed successively with saturated aq NH₄Cl and brine, dried (MgSO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (hexane/ EtOAc=5.5:1) to give bis-trichloroacetimidate 9 (301.7 mg, 97%) as a colorless solid. Analytical sample (colorless needles) was obtained by recrystallization from hexane; mp 71.5–74.5 °C; $[\alpha]_D^{26.1}$ +1.07 (c 0.655, CHCl₃); ν_{max} (KBr) 3336, 2989, 1718, 1658, 1313, 1227, 1167, 1076, 1059, 1009, 872, 800, 650, 571 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 8.45 (br s, 1H), 8.37 (br s, 1H), 6.09–6.05 (m, 1H), 5.51–5.40 (m, 2H), 4.17 (dd, J=7.2, 10.9 Hz, 1H), 4.14 (dd, J=7.1, 10.9 Hz, 1H), 2.27 (d, J=1.5 Hz, 3H), 1.41 (d, J=6.2 Hz, 3H), 1.27 (t, J=7.2 Hz, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 166.0, 161.8, 161.3, 150.9, 119.3, 91.3, 91.0, 82.7, 74.2, 60.0, 16.0, 15.2, 14.2. Anal. Calcd for C₁₃H₁₆Cl₆N₂O₄: C, 32.73; H, 3.38; N, 5.87. Found: C, 32.74; H, 3.41; N, 5.82%.

4.8.3. Ethyl 2-((4R,5R)-5-((R)-1-(trichloroacetimidoyloxy)ethyl)-2-(trichloromethyl)-4,5-dihydro-4-methyloxazol-4-yl)acetate (10) and ethyl 2-((4S,5R)-5-((R)-1-(trichloroacetimidoyloxy)ethyl)-2-(trichloromethyl)-4,5-dihydro-4methyloxazol-4-yl)acetate (11)

To a solution of the trichloroacetimidate **9** (517.9 mg, 1.09 mmol) in THF (44 mL) was added *tert*-BuOK (99.8 mg, 0.889 mmol) in one portion under N₂ atmosphere at below -90 °C, and the mixture was stirred for ca. 2 h. The reaction mixture was poured into cold saturated aq NH₄Cl and extracted with EtOAc. The extract was washed with brine, dried (MgSO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (hexane/EtOAc=5:1) to give *cis*-oxazoline **10** (less polar, 277.1 mg, 54%) and *trans*-oxazoline **11** (more polar, 189.9 mg, 37%). Analytical sample of **10** and **11** were obtained by recrystallization from EtOAc/hexane.

4.8.3.1. cis-Oxazoline **10**. Colorless needles; mp 95.3– 97.5 °C; $[\alpha]_D^{27.4}$ –104 (c 0.575, CHCl₃); ν_{max} (KBr) 3338, 2989, 1718, 1660, 1338, 1306, 1228, 1201, 1119, 1080, 796, 687, 652 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 8.43 (br s, 1H), 5.45 (dq, *J*=0.9, 6.4 Hz, 1H), 4.75 (d, *J*=0.9 Hz, 1H), 4.19 (dd, *J*=7.2, 10.8 Hz, 1H), 4.17 (dd, *J*=7.1, 10.9 Hz, 1H), 2.98 (d, *J*=17.7 Hz, 1H), 2.85 (d, *J*=18.2 Hz, 1H), 1.46 (d, *J*=6.4 Hz, 3H), 1.44 (d, *J*=0.6 Hz, 3H), 1.28 (t, *J*=7.1 Hz, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 171.4, 161.1, 160.8, 92.0, 91.0, 86.7, 73.9, 70.4, 61.0, 39.8, 26.5, 15.3, 14.1. Anal. Calcd for C₁₃H₁₆Cl₆N₂O₄: C, 32.73; H, 3.38; N, 5.87. Found: C, 33.02; H, 3.38; N, 5.81%. 4.8.3.2. trans-Oxazoline **11**. Colorless needles; mp 69.5– 70.3 °C; $[\alpha]_D^{27.9}$ -66.5 (c 0.580, CHCl₃); ν_{max} (KBr) 3327, 2991, 1718, 1668, 1331, 1284, 1248, 1109, 1072, 1028, 796, 677, 650 cm⁻¹; ¹H NMR (270 MHz, CDCl₃) δ 8.45 (br s, 1H), 5.34 (dq, *J*=1.1, 6.4 Hz, 1H), 5.07 (d, *J*=1.1 Hz, 1H), 4.17 (q, *J*=7.1 Hz, 2H), 2.69 (d, *J*=15.6 Hz, 1H), 2.59 (d, *J*=15.6 Hz, 1H), 1.502 (s, 3H), 1.497 (d, *J*=6.4 Hz, 3H), 1.28 (t, *J*=7.2 Hz, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 170.1, 161.4, 160.9, 91.1, 90.0, 86.5, 73.1, 71.0, 60.9, 45.5, 19.8, 15.8, 14.2. Anal. Calcd for C₁₃H₁₆Cl₆N₂O₄: C, 32.73; H, 3.38; N, 5.87. Found: C, 32.95; H, 3.37; N, 5.87%.

4.8.4. (3R,4R,5R)-3-Benzamido-5-hydroxy-3-methyl-4hexanolide (12)

To a solution of the *cis*-oxazoline **10** (586.1 mg, 1.23 mmol) in EtOH (21 mL) was added 3 M HCl (21 mL) and the mixture was refluxed for ca. 19 h. The reaction mixture was concentrated in vacuo and added acetone (9 mL) and saturated aq NaHCO₃ (22 mL). To the mixture was added dropwise benzovl chloride (428 µL, 3.68 mmol), and the mixture was stirred for ca. 1 h at rt. The mixture was concentrated in vacuo and the residue was diluted with brine and the whole mixture was extracted with EtOAc. The extract was dried (MgSO₄) and concentrated in vacuo. The residue was purified by flash column chromatography (hexane/EtOAc=1:2.5) to give lactone 12 (275.0 mg, 85%). Analytical sample (colorless rod) was obtained by recrystallization from EtOAc/hexane; mp 173.0–174.5 °C; $[\alpha]_D^{26.6}$ +46.3 (c 0.580, EtOH); ν_{max} (KBr) 3452, 3346, 3309, 2987, 1780, 1759, 1666, 1635, 1541, 1522, 1315, 1292, 1194, 1142, 989, 725, 694 cm^{-1} ; ¹H NMR (270 MHz, CDCl₃) δ 7.77–7.68 (m, 2H), 7.57-7.40 (m, 3H), 6.14 (br s, 1H), 4.84 (d, J=1.9 Hz, 1H), 4.22 (ddq, J=1.9, 6.4, 6.4 Hz, 1H), 2.94 (d, J=17.5 Hz, 1H), 2.88 (d, J=17.5 Hz, 1H), 1.78 (s, 3H), 1.54 (d, J=6.2 Hz, 3H), 1.38 (d, J=6.4 Hz, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 175.2, 167.5, 134.1, 132.0, 128.7, 126.8, 87.7, 66.3, 59.7, 42.5, 20.7, 20.3. Anal. Calcd for C₁₄H₁₇NO₄: C, 63.87; H, 6.51; N, 5.32. Found: C, 63.74; H, 6.51; N, 5.27%.

4.8.5. (3S,4R,5R)-3-Benzamido-5-hydroxy-3-methyl-4hexanolide (13)

To a solution of the *trans*-oxazoline **11** (111.7 mg, 0.234 mmol) in EtOH (5 mL) was added 3 M HCl (5 mL) and the mixture was refluxed for ca. 15 h. The reaction mixture was concentrated in vacuo and added acetone (1.65 mL) and saturated aq NaHCO₃ (4.2 mL). To the mixture was added dropwise benzoyl chloride (81.5 µL, 0.702 mmmol), and the mixture was stirred for ca. 1 h at rt. The mixture was concentrated in vacuo and the residue was diluted with brine and the whole mixture was extracted with EtOAc. The extract was dried (MgSO₄) and concentrated in vacuo. The residue was purified by flash column chromatography (hexane/ EtOAc=1:2.5) to give lactone 13 (50.8 mg, 82%). Analytical sample (colorless rod) was obtained by recrystallization from EtOAc/hexane; mp 124.5–126.0 °C; $[\alpha]_D^{27.0}$ –37.7 (c 0.560, EtOH); v_{max} (KBr) 3321, 2976, 1782, 1645, 1541, 1300, 1159, 1138, 985, 729 cm⁻¹; ¹H NMR (270 MHz,

CDCl₃) δ 8.59 (br s, 1H), 7.80–7.72 (m, 2H), 7.54–7.37 (m, 3H), 4.31 (dq, *J*=6.7, 9.5 Hz, 1H), 4.09 (s, 1H), 3.59 (d, *J*=17.5 Hz, 1H), 2.62 (d, *J*=17.5 Hz, 1H), 2.24 (d, *J*=9.8 Hz, 1H), 1.74 (s, 3H), 1.51 (d, *J*=6.6 Hz, 3H); ¹³C NMR (67.8 MHz, CDCl₃) δ 174.6, 167.3, 134.2, 131.8, 128.6, 126.9, 87.4, 65.1, 59.9, 41.8, 22.3 (2). Anal. Calcd for C₁₄H₁₇NO₄: C, 63.87; H, 6.51; N, 5.32. Found: C, 63.76; H, 6.52; N, 5.27%.

4.8.6. 3-Benzamido-2,3,6-trideoxy-3-C-methyl-D-lyxohexopyranose (N-Bz-D-vancosamine)

To a solution of the lactone 12 (75.6 mg, 0.287 mmol) in dry THF (4.2 mL) was added dropwise DIBAL (1.46 mL, 0.98 M in hexane, 1.43 mmol) under Ar atmosphere at -60 °C, and the reaction mixture was stirred for 1.5 h at -60° to -50° C. The reaction was quenched by the dropwise addition of acetone/MeOH (1:1, 3 mL) and the mixture was allowed to warm gradually to rt with stirring for ca. 4 h. The mixture was filtered through a pad of Celite. The filter cake was washed with acetone/MeOH (1:1) and then the filtrate was concentrated in vacuo. The residue was purified by flash column chromatography (EtOAc) to give N-Bz-D-vancosamine (39.9 mg, 52%) along with over-reduced triol (11.8 mg, 15%). Analytical sample (colorless rod) was obtained by recrystallization from EtOAc/hexane; mp 152.0-154.0 °C (lit.²⁰ 155 °C); $[\alpha]_{D}^{29.1}$ +81.9 (after 10 min), +34.7 (after 1 day, constant) (c 0.525, MeOH) (lit.²⁰ L-form $[\alpha]_D^{20}$ -86.6 (c 0.5, MeOH)); v_{max} (KBr) 3332, 2931, 1645, 1581, 1533, 1491, 1284, 1097, 1016, 710 cm⁻¹; ¹H NMR (270 MHz, DMSO d_6 , after 1 month) data were in agreement with the reported in lit. 20; ¹³C NMR (67.8 MHz, DMSO- d_6) δ α -isomer 165.6, 135.7, 130.9, 128.1, 126.9, 90.1, 71.2, 62.6, 54.3, 35.2, 23.2, 17.47; β-isomer 165.7, 135.6, 130.8, 128.0, 127.0, 92.0, 70.0, 68.2, 56.1, 38.7, 21.4, 17.52. Anal. Calcd for C14H19NO4: C, 63.38; H, 7.22; N, 5.28. Found: C, 63.21; H, 7.20; N, 5.24%.

4.8.7. 3-Benzamido-2,3,6-trideoxy-3-C-methyl-D-xylohexopyranose (N-Bz-3-epi-D-vancosamine)

To a solution of the lactone 13 (33.1 mg, 0.126 mmol) in dry THF (1.9 mL) was added dropwise DIBAL (641 µL, 0.98 M in hexane, 0.628 mmol) under Ar atmosphere at -60 °C, and the reaction mixture was stirred for 1 h at -60° to -55 °C. The reaction was quenched by the dropwise addition of acetone/MeOH (1:1, 2 mL) and the mixture was allowed to warm gradually to rt with stirring for ca. 4 h. The mixture was filtered through a pad of Celite. The filter cake was washed with acetone/MeOH (1:1), and then the filtrate was concentrated in vacuo. The residue was purified by flash column chromatography (EtOAc) to give N-Bz-3-epi-D-vancosamine (24.1 mg, 72%). Analytical sample (colorless solid) was obtained by recrystallization from EtOAc/hexane; mp 222.0–224.0 °C (lit.²⁰ 230–233 °C); $[\alpha]_D^{28.0}$ +69.6 (after 10 min), $[\alpha]_D^{29.7}$ +45.7 (after 1 day, constant) (*c* 0.505, MeOH) (lit.²⁰ L-form $[\alpha]_D^{20}$ -61.3 (*c* 0.5, MeOH)); ν_{max} (KBr) 3361, 3239, 2979, 1643, 1549, 1362, 1327, 1261, 1113, 1099, 1016, 964, 868, 802, 723, 694, 631 cm⁻¹; ¹H NMR (270 MHz, DMSO- d_6 , after 1 month) data were in agreement with the reported in lit. 20; ¹³C NMR (67.8 MHz, DMSO- d_6) δ α-isomer 164.9, 135.4, 131.1, 128.5, 126.3, 90.5, 67.6, 62.5, 54.7, 35.4, 22.8, 17.34; β-isomer 166.7, 135.8, 130.8, 127.9, 127.3, 92.1, 68.4, 67.9, 57.1, 37.0, 23.3, 17.31. Anal. Calcd for C₁₄H₁₉NO₄: C, 63.38; H, 7.22; N, 5.28. Found: C, 63.18; H, 7.23; N, 5.23%.

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References and notes

- 1. Weymouth-Wilson, A. C. Nat. Prod. Rep. 1997, 14, 99-110.
- Paloma, L. G.; Smith, J. A.; Chazin, W. J.; Nicolaou, K. C. J. Am. Chem. Soc. 1994, 116, 3697–3708.
- 3. Nedal, A.; Zotchev, S. B. Appl. Microbiol. Biotechnol. 2004, 64, 7-15.
- Yu, T.-W.; Müller, R.; Müller, M.; Zhang, X.; Draeger, G.; Kim, C.-G.; Leistner, E.; Floss, H. G. J. Biol. Chem. 2001, 276, 12546–12555.
- 5. Matsushima, Y.; Kino, J. Tetrahedron Lett. 2005, 46, 8609-8612.
- 6. Matsushima, Y.; Kino, J. Tetrahedron Lett. 2006, 47, 8777-8780.
- 7. Bergmeier, S. C. Tetrahedron 2000, 56, 2561-2576.
- Kochi, T.; Tang, T. P.; Ellman, J. A. J. Am. Chem. Soc. 2002, 124, 6518– 6519; J. Am. Chem. Soc. 2003, 125, 11276–11282 and references therein.
- A few examples of intramolecular conjugate additions of trichloroacetimidates, which were prepared in situ, in a cyclic system were reported: (a) For α,β-unsaturated ester, see: Brown, J. R.; Nishimura, Y.; Esko, J. D. *Bioorg. Med. Chem. Lett.* 2006, *16*, 532–536; (b) For α,β-unsaturated nitrile, see: Fraser-Reid, B.; Burgey, C. S.; Vollerthum, R. *Pure Appl. Chem.* 1998, *70*, 285–288; (c) For α,β-unsaturated sulfones, see: Li, X. C.; Fuchs, P. L. *Synlett* 1994, 629–630.
- There are some useful reactions involving the trichloroacetimidates for the introduction of a nitrogen functionality. (a) For electrophile-promoted intramolecular aminations of trichloroacetimidates derived from allylic and homoallylic alcohols, see: Lee, H.-S.; Kang, S. H. Synlett 2004, 1673–1685 and references therein, (b) For acid-promoted intramolecular epoxide opening of trichloroacetimidates, see: Bernet, B.; Vasella, A. *Tetrahedron Lett.* 1983, 24, 5491–5494; Schmidt, U.; Respondek, M.; Lieberknecht, A.; Werner, J.; Fischer, O. Synthesis 1989, 256–261; Hatakeyama, S.; Matsumoto, H.; Fukuyama, H.; Mukugin, Y.; Irie, H. *J. Org. Chem.* 1997, 62, 2275–2279; Matsushima, Y.; Nakayama, T.; Tohyama, S.; Eguchi, T.; Kakinuma, K. *J. Chem. Soc., Perkin Trans. 1* 2001, 569–577 and see also: Rondot, C.; Retailleau, P.; Zhu, J. Org. *Lett.* 2007, 9, 247–250 and references therein; (c) For Overman rearrangement, see: Overman, L. E. Acc. Chem. Res. 1980, 13, 218–224.
- Starting allylic alcohols were, respectively, prepared according to the literatures below: (a) Trost, B. M.; Suriveta, J.-P.; Toste, F. D. J. Am. Chem. Soc. 2004, 126, 11592–11602; (b) lit. 5; (c) lit. 6; (d) Xu, D.; Crispino, G. A.; Sharpless, K. B. J. Am. Chem. Soc. 1992, 114, 7570–7571; Becker, H.; Soler, M. A.; Sharpless, K. B. Tetrahedron 1995, 51, 1345–1376.
- 12. During the imidate formation a trace amount of conjugate adducts was obtained at low temperature and prolonged time or increased temperature afforded the products, but only in low yield.
- It was reported that in an oxazoline ring the small coupling constant (ca. 6 Hz) corresponded to trans form and the large one (ca. 9 Hz) to cis form; see: Roush, W. R.; Straub, J. A.; Brown, R. J. J. Org. Chem. 1987, 52, 5127–5136.

- 14. Hirama, M.; Shigemoto, T.; Itô, S. *Tetrahedron Lett.* **1985**, *26*, 4133–4136, 4137–4140, and references therein.
- Starting homoallylic alcohols were, respectively, prepared according to the literatures below: (a) Garaas, S. D.; Hunter, T. J.; O'Doherty, G. A. J. Org. Chem. 2002, 67, 2682–2685; (b) Moinuddin, M.; O'Doherty, G. A. Carbohydr. Res. 2006, 341, 1505–1521; (c) lit. 5; (d) Uchida, K.; Ishigami, K.; Watanabe, H.; Kitahara, T. Tetrahedron 2007, 63, 1281–1287.
- 16. For example, the irradiation of H-4' on the *cis*-dihydrooxazine **5a** yielded an NOE of ca. 6% on H-6', and this result was supported by their trans-diaxial large coupling constants $(J_{\text{H-4',H-5'ax}} \text{ and } J_{\text{H-5'ax,H-6'}}=\text{ca.}$ 11 Hz) of **5a**.
- Syntheses of vancosamine derivatives, see: Hsu, D.-S.; Matsumoto, T.; Suzuki, K. Synlett 2006, 469–471; Trost, B. M.; Jiang, C.; Hammer, K. Synthesis 2005, 3335–3345 and references therein.
- The known acetonide 7 was prepared from L-threonine, see: Aoyagi, S.; Wang, T.-C.; Kibayashi, C. J. Am. Chem. Soc. 1993, 115, 11393–11409 and references therein.
- 19. The irradiation of methyne hydrogen (H-5') on the *cis*-oxazoline **10** yielded no NOE on the methylene protons (H-2). On the contrary, the irradiation of the hydrogen corresponding to the *trans*-oxazoline **11** yielded an NOE of ca. 4% on the one methylene proton (H-2).
- Fronza, G.; Fuganti, C.; Grasselli, P.; Pedrocchi-Fantoni, G. *Tetrahedron* Lett. 1981, 22, 5073–5076; J. Carbohydr. Chem. 1983, 2, 225–248.